

## SURGERY

# Contribution of Lumbar Spine Pathology and Age to Paraspinal Muscle Size and Fatty Infiltration

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**Study Design.** Retrospective chart analysis of 199 individuals aged 18 to 80 years scheduled for lumbar spine surgery.

**Objective.** The purpose of this study was to quantify changes in muscle cross-sectional area (CSA) and fat signal fraction (FSF) with age in men and women with lumbar spine pathology and compare them to published normative data.

**Summary of Background Data.** Pathological changes in lumbar paraspinal muscle are often confounded by age-related decline in muscle size (CSA) and quality (fatty infiltration). Individuals with pathology have been shown to have decreased CSA and fatty infiltration of both the multifidus and erector spinae muscles, but the magnitude of these changes in the context of normal aging is unknown.

**Methods.** Individuals aged 18 to 80 years who were scheduled for lumbar surgery for diagnoses associated with lumbar spine pain or pathology were included. Muscle CSA and FSF of the multifidus and erector spinae were measured from preoperative T2-weighted magnetic resonance images at the L4 level. Univariate and multiple linear regression analyses were performed for each outcome using age and sex as predictor variables. Statistical comparisons of univariate regression parameters (slope and intercept) to published normative data were also performed.

**Results.** There was no change in CSA with age in either sex ( $P > 0.05$ ), but women had lower CSAs than men in both muscles ( $P < 0.0001$ ). There was an increase in FSF with age in erector spinae and multifidus muscles in both sexes

( $P < 0.0001$ ). Multifidus FSF values were higher in women with lumbar spine pathology than published values for healthy controls ( $P = 0.03$ ), and slopes tended to be steeper with pathology for both muscles in women ( $P < 0.08$ ) but not in men ( $P > 0.31$ ).

**Conclusion.** Lumbar muscle fat content, but not CSA, changes with age in individuals with pathology. In women, this increase is more profound than age-related increases in healthy individuals.

**Key words:** age, cross sectional area, fatty infiltration, lumbar spine pathology, magnetic resonance imaging.

**Level of Evidence:** 3  
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Low back pain (LBP) is highly prevalent in the United States, with approximately 65% to 85% of the general population experiencing some low back pain within their lifetime.<sup>1</sup> Although most LBP is considered self-limiting in nature, recent evidence suggests that a high proportion of individuals develop recurrent symptoms, leading to poor functional outcomes and increased health care utilization.<sup>1,2</sup> Changes in muscle size and fat content are often associated with LBP symptoms and lumbar spine pathology<sup>3,4</sup>; however, the magnitude of these pathological changes is confounded by natural age-related changes such as sarcopenia, fatty infiltration, and decreased torque production.<sup>5</sup> Muscle physiological cross-sectional area is commonly used as an indicator for muscle force production capacity,<sup>6</sup> and cross-sectional area (CSA) has been quantified using magnetic resonance imaging (MRI) region of interest segmentation analyses.<sup>7</sup> Measures of total muscle size or volume in isolation do not take into account fatty infiltration, which is associated with aging and pathology.<sup>8</sup> Since fatty infiltration in muscle decreases the proportion of contractile tissue capable of producing force, it is important to understand how muscle size and fatty infiltration change, both with age, and in the presence of pathology. Understanding muscle loss in the presence of lumbar spine pathology requires an understanding of how muscle loss occurs with aging. If pathology yields unique rates of muscle loss,

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new strategies for resolving muscle loss should be a clinical goal for functional improvement, as standard exercise strategies do not appear to reverse these changes in this population.<sup>9,10–12</sup>

Previous literature suggests that LBP and pathology alter CSA and fatty infiltration, with most studies demonstrating decreases in muscle size and increases in fatty infiltration in those with symptoms compared to their healthy counterparts.<sup>13–15</sup> However, the extent of these morphological changes is unclear when considering physiological declines observed with normal aging.<sup>16–18</sup> Effects of aging and pathology on muscle size and quality have been observed in other musculoskeletal regions, such as the rotator cuff and thigh muscles. In the rotator cuff, there is evidence that muscle size decreases more with age in individuals with tears compared to those with no known pathology.<sup>19</sup> Similarly, fatty infiltration seems to be more pronounced in individuals with rotator cuff tears compared to healthy controls across a spectrum of ages.<sup>20</sup> There is also evidence of age-related decreases in CSA and increases in fatty infiltrate in the thigh muscles, although no comparisons have been made to pathological conditions in this muscle region.<sup>5,21</sup> Currently, there is no information on the differential effect of age and pathology on changes in the lumbar musculature. The purpose of this study was to determine the relationship between age and muscle CSA and fatty infiltrate in men and women with LBP or pathology who were scheduled to undergo lumbar spine surgery, and compare these relationships to published normative data on fatty infiltrate in healthy individuals. The overall goal of this work is to begin to uncouple the effects of pain/injury and age on atrophic changes in lumbar spine musculature. We hypothesize that muscle size and quality will decline with age, and this decline will be more pronounced in patients with lumbar spine pathology.

## MATERIALS AND METHODS

### Study Participants

MRIs from 236 patients were screened based on an initial chart query using current procedural terminology (CPT) codes for lumbar spine surgical procedures from 2005 to 2015 at the University of California San Diego hospital database. Individuals were included in this screen if they were between 18 and 80 years of age, and were undergoing a surgical procedure of the lumbar spine. Patients were excluded from the analyses if they did not have a concurrent diagnosis (ICD-9) or procedural code associated with lumbar spine-related symptoms, or did not have imaging of the lumbar spine. Surgical and diagnosis codes included in the query and initial screening are listed in Table 1. From the initial 236 patients queried, 37 patients were excluded from the analyses; 17 patients were excluded because of existence of instrumentation at the L4 level affecting MR signal intensity analyses, 7 were excluded because of CPT codes unrelated to LBP or degenerative pathology (*i.e.*, skin mass, neoplasm), and 9 were excluded because of analytical

limitations (*i.e.*, T2 images not available, motion artifact), and 4 were excluded because of acute trauma with lumbar vertebral fracture from a motor vehicle accident. After screening and exclusions, a total of 199 patients were retained for analysis (Figure 1). This study was approved by the local institutional review board (IRB #071983).

### MRI Acquisition and Measurements

Regions of interest from T2-weighted axial magnetic resonance images taken from a single slice estimated to be closest to the midlevel of the L4 vertebra were used to measure muscle CSA and fat content with custom written Matlab software (Mathworks, Natick MA). For CSA measurements, regions of interest for both multifidus and erector spinae muscles were seeded and segmented bilaterally using Osirix software,<sup>22</sup> based on fascial plane separations using the facet joint as a landmark between the multifidus and erector spinae, and the lumbosacral fascia posteriorly<sup>23</sup> (Figure 2A). Pixels were identified as either fat or muscle by fitting a two-term Gaussian model to the histogram of pixel intensities from segmented regions of interest, and finding the intersection of the Gaussian distributions (Figure 2B). Pixel values above the intersection were classified as fat, and pixels below were classified as muscle. Fat content was measured using the fat signal fraction (FSF) and was calculated with the following equation<sup>24</sup>:

$$FSF = \frac{npixels_{fat}}{npixels_{fat} + npixels_{muscle}}$$

### Statistical Analysis

Measures of CSA and FSF were averaged between left and right muscle regions of interest for the multifidus and erector spinae muscles separately. Age was analyzed as a continuous variable, and sex was analyzed as a dichotomous variable. Differences in demographic characteristics between sexes were analyzed using independent *t* tests for continuous variables, and  $\chi^2$  tests for categorical variables. Separate linear regression analyses were performed for CSA and FSF measurements with both age and sex as predictor variables in a single model. Univariate regressions between age and CSA or FSF were then performed for each sex for comparison to data from a healthy cohort. Parameter estimates and intercepts from the univariate regression analyses were compared to healthy subjects from Crawford *et al*<sup>25</sup> using independent *t* tests (personal correspondence). All statistical analyses were performed using SAS software (SAS 9.3, SAS Institute, Cary, NC). Statistical significance was set at  $P < 0.05$  and trends were defined as  $P < 0.1$ .

## RESULTS

Mean (SD) patient age was 58.7 (13.5) years with no significant differences in age between men and women ( $P = 0.79$ ). There were a larger proportion of women ( $n = 116$ , 58.2%) compared to men ( $n = 83$ , 41.8%). The most common preoperative lumbar diagnoses were

**TABLE 1. Surgical and Diagnostic Codes Used for Inclusion Criteria in Chart Queries**

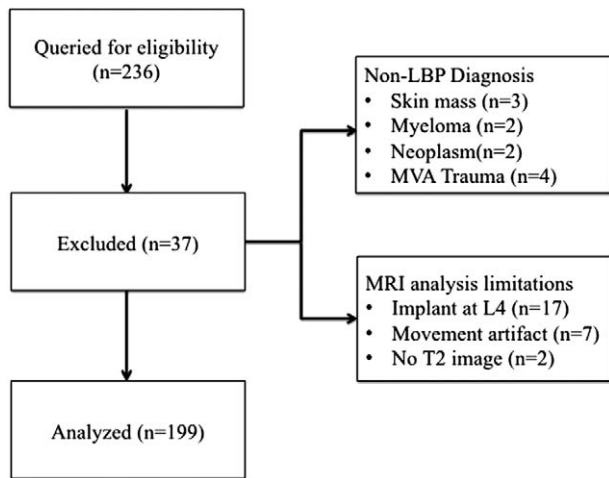
Surgical Codes	Description	ICD-9 Code	Diagnosis
22585	Fusion of additional interspaces	719.45	Pain in joint, pelvic region and thigh
22612	Posterolateral lumbar fusion	721.3	Lumbosacral spondylosis without myelopathy
22614	Posterolateral lumbar fusion, additional segments	722.52	Degeneration of lumbar or lumbosacral intervertebral disc
22630	Posterior interbody fusion, lumbar	724.02	Spinal stenosis, lumbar region, without neurogenic claudication
22633	Combined fusion, posterolateral fusion, with posterior interbody fusion	724.03	Spinal stenosis, lumbar region, with neurogenic claudication
22634	Combined fusion, posterolateral fusion, with posterior interbody fusion (each additional interspace/segment)	724.2	Lumbago
63005	Laminectomy without facetectomy, foraminotomy or discectomy, lumbar, except for spondylolisthesis	724.4	Sciatica
63012	Laminectomy with removal of abnormal facets and/or pars interarticularis with decompression, for spondylolisthesis, lumbar	724.5	Thoracic or lumbosacral neuritis or radiculitis, unspecified
63017	Laminectomy without facetectomy, foraminotomy or discectomy, lumbar, except for spondylolisthesis (>two vertebral segments)	729.5	Backache, unspecified
63030	Laminotomy (hemilaminectomy), including partial facetectomy, foraminotomy and/or excision of herniated disc, lumbar	782	Backache, unspecified
63035	Laminotomy (hemilaminectomy), including partial facetectomy, foraminotomy and/or excision of herniated disc, lumbar (each additional interspace)		
63042	Laminotomy (hemilaminectomy), including partial facetectomy, foraminotomy and/or excision of herniated disc, reexploration, lumbar		
60344	Laminotomy (hemilaminectomy), including partial facetectomy, foraminotomy and/or excision of herniated disc, reexploration, lumbar (each additional interspace)		
63047	Laminectomy, facetectomy and foraminotomy, lumbar		

radiculopathy (42.2%), nonspecific LBP (26.1%), and spinal stenosis (10.6%). All other related diagnoses were categorized as “other” and accounted for 21.1% of the cases. There were no differences in types of diagnoses between sexes (Table 2). Similarly, there were no differences in age across the diagnostic categories ( $P = 0.35$ ). Linear regression model results demonstrated no effect of age on muscle CSA ( $\beta = -0.0002$  and  $-0.0118$  for multifidus and erector spinae respectively,  $P > 0.59$ ), but a significant sex effect for multifidus ( $\beta = -2.26$ ,  $P < 0.0001$ ) and erector spinae muscle CSA ( $\beta = -3.19$ ,  $P < 0.0001$ ) wherein men displayed a larger CSA for both spine muscles (Figure 3C and D). Age and sex were significant predictors of FSF in the multifidus (age:  $\beta = 0.004$ , sex:  $\beta = 0.0600$ ;  $P < 0.0001$ ), whereas only age predicted FSF in the erector spinae (age:  $\beta = 0.0040$ ,  $P < 0.0001$ , sex:  $\beta = 0.0201$ ;  $P = 0.15$ ) muscle

(Figure 3A and B). Women displayed more FSF than men in the multifidus, and older ages were associated with higher FSF in both muscles.

In the univariate sex stratified analyses, none of the variance in CSA was explained by age in either sex or muscle group ( $r^2 < 0.003$ ,  $P > 0.84$ ). For FSF, age accounted for approximately 33% of the variance in FSF in the multifidus ( $P < 0.0001$ ), and 32% in the erector spinae in women ( $P < 0.0001$ ). In men, age accounted for 22% of the variance in FSF in the multifidus ( $P < 0.0001$ ) and 14% in the erector spinae ( $P = 0.0005$ ). In a subanalysis, we compared the sex-stratified univariate regression estimates between age and FSF from our study with a previous study that reported normative values for healthy individuals aged 20 to 62 years.<sup>25</sup> There was a trend toward a steeper slope in women with pathology compared to women without pathology for





**Figure 1.** Flow diagram of participant screening, exclusion, and analysis.

both multifidus ( $P=0.08$ ) and erector spinae muscles ( $P=0.08$ ), such that women with pathology displayed more age-related increases in FSF (Figure 4B and D). For men, there were no significant differences in slopes between healthy and pathological populations ( $P>0.28$ ). When comparing intercepts between healthy and pathological populations, men with pathology tended to display higher intercepts in both multifidus ( $P=0.06$ ) and erector spinae muscles ( $P=0.05$ ) (Figure 4A and C), and women with pathology had a higher intercept only in the multifidus ( $P=0.03$ ). Compared to a healthy population, men with pathology display overall more FSF in both multifidus and erector spinae muscles, and women display more FSF in the multifidus (Table 3). Recognizing that differences in age range between the population in the current study and the healthy cohort may affect the results, we also performed subcomparisons using only patients younger than 65 years ( $N=129$ ), yielding similar main effects of age and sex. However, trends observed between intercepts for healthy and pathological groups in the erector spinae were lost ( $P<0.17$ ), as well as the difference in intercept between healthy and pathological women for the multifidus ( $P=0.29$ ). The trend for a difference between intercepts

for multifidus FSF in men became significant ( $P=0.04$ ). All other results remained consistent with the full cohort comparison.

Owing to differences in analytical methods for measuring muscle size, comparisons between normal and pathological trajectories for CSA were not performed (Figure 4).

## DISCUSSION

The results of this study demonstrate that there is no significant effect of age on changes in CSA of either multifidus or erector spinae muscles in individuals with lumbar spine pathology; however, there are significant increases in paraspinal muscle FSF with age in both sexes. When compared to previous published data on healthy individuals across a smaller age range, overall levels of FSF are higher in individuals with pathology across all ages. Additionally, a trend toward a more pronounced increase in FSF with age is seen in women with pathology, but not in men. Finally, men with pathology tend to display higher FSF values compared to a healthy population for both muscles and women with pathology tend to have higher FSF values in the multifidus only. When limiting comparisons to include only those younger than 65 years, men displayed higher FSF values for the multifidus only, and women did not display higher FSF values in either muscle when compared to the healthy cohort, although the trends toward a more pronounced increase in FSF with age in women were retained for both muscles.

## CSA and FSF in Healthy and Pathological Populations

Direct comparisons of CSA values from the current study to previous literature are limited by variation in segmentation methodology, such as considering the erector spinae and multifidus muscles as one unit, as well as data-reporting methods such as normalization to vertebral body size, which we did not measure in this study. One study using similar segmentation methods in patients with LBP ranging from 18 to 60 years reported average CSA values of 10.1 (1.5)  $\text{cm}^2$  and 18.5 (3.9)  $\text{cm}^2$  at the L4 level for the multifidus and erector spinae muscles respectively, which is consistent with the data from the current study.<sup>7</sup> To our knowledge, no data exist on CSA values in healthy individuals that are

**Figure 2.** Magnetic resonance image segmentation for CSA and FSF analysis. (A) Regions of interest for multifidus (M) and erector spinae (E) muscles, bordered anteriorly by the lamina and spinous process, and posteriorly by the lumbosacral fascia. (B) Representative histogram of pixel intensities across a spectrum of fat and muscle. Thresholds were defined on a patient-by-patient basis as the intersection between the Gaussian distribution of fat and water (x). CSA, cross-sectional area; FSF, fat signal fraction.

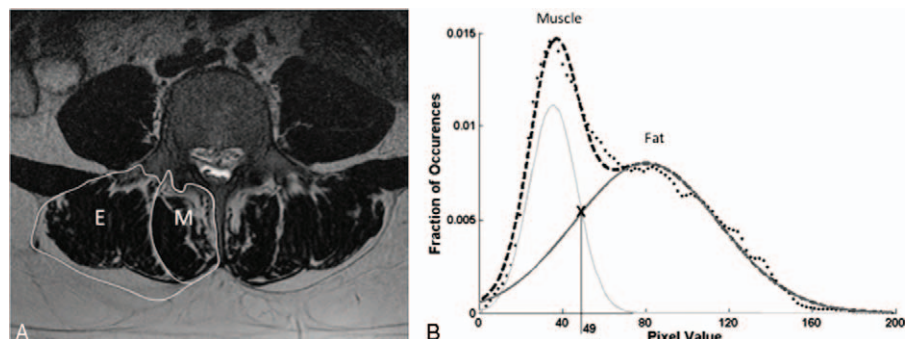
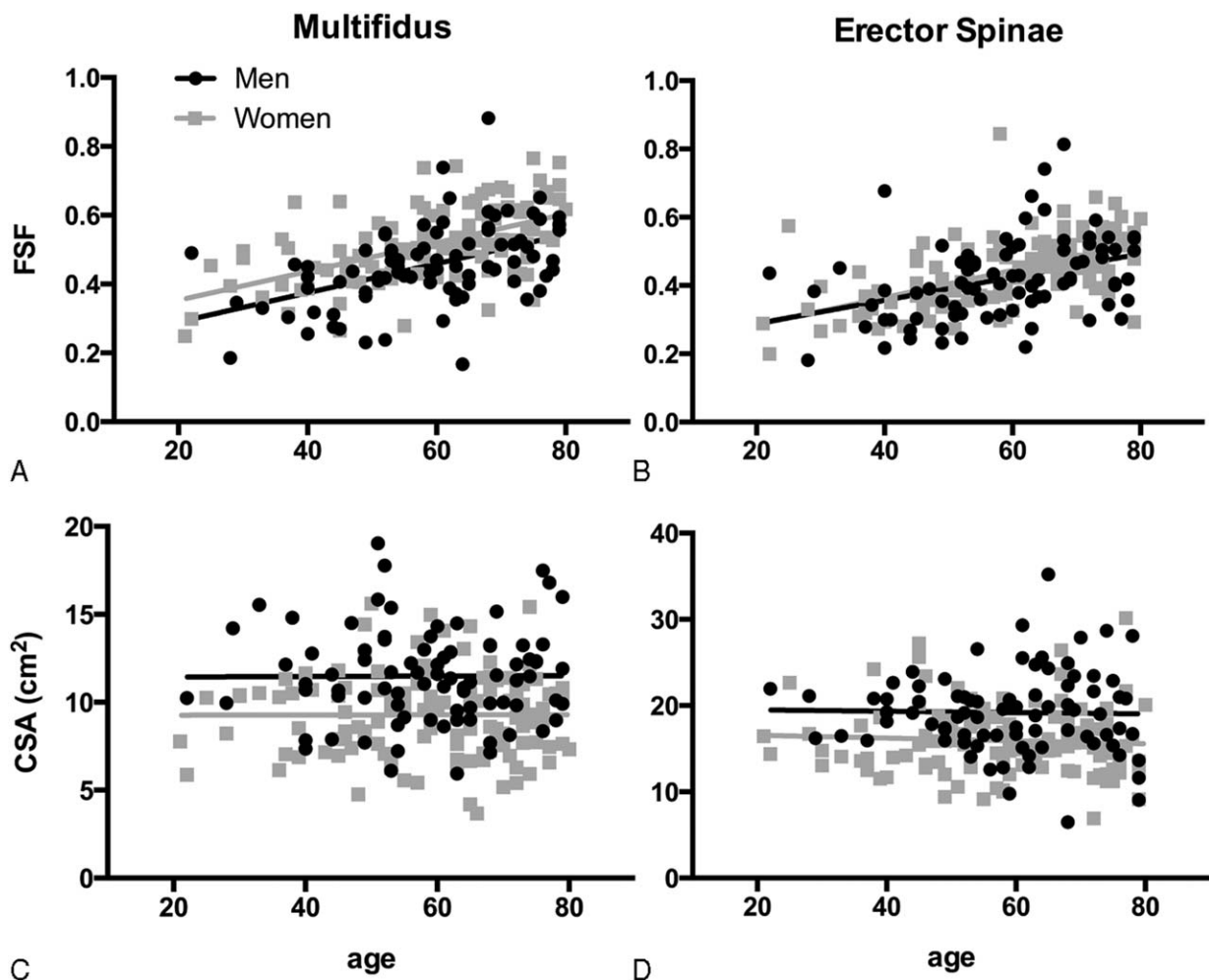


TABLE 2. Patient Demographic Characteristics			
	Female (n = 116)	Male (n = 83)	P
Patients (%) n = 199	59.1	40.9	0.011
Age, y (SD)	58.6 (13.3)	59.1 (14.2)	0.79
Diagnosis (% of total)			
Nonspecific LBP	15.3	10.8	0.89
Radiculopathy	25.6	17.2	0.88
Stenosis	4.9	5.4	0.82
Other	13.3	7.4	0.86
CSA, cm <sup>2</sup>			
Multifidus	9.29 (2.38)	11.49 (2.69)	<0.0001
Erector spinae	15.92 (3.96)	19.20 (4.70)	<0.0001
FSF			
Multifidus	0.52 (0.11)	0.45 (0.12)	<0.0001
Erector Spinae	0.44 (0.10)	0.42 (0.12)	0.17

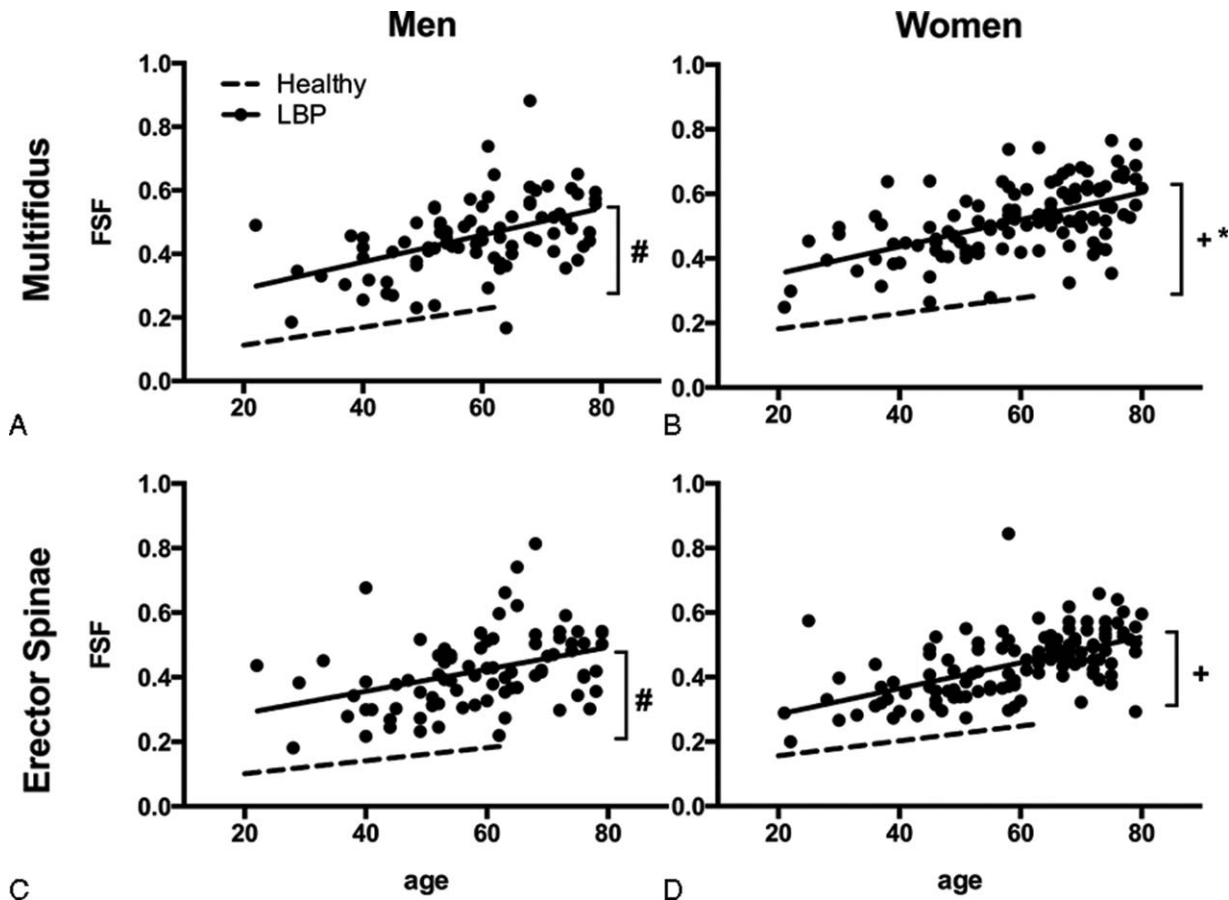
CSA, cross-sectional area; FSF, fat signal fraction; LBP, low back pain.

methodologically consistent with and directly comparable to values from the current study. When comparing levels of fatty infiltrate in the lumbar multifidus for individuals with and

without LBP, Fischer *et al*<sup>26</sup> reported mean FSF percentage values of 20.1% (range 4.3%–73.4%) in individuals with chronic LBP symptoms, and Crawford *et al*<sup>25</sup> reported mean



**Figure 3.** Regression plots for men (black) and women (grey) for multifidus (FSF in Panel A, CSA in Panel C) and erector spinae (FSF in Panel B, CSA in Panel D). There was a significant effect of sex in both FSF and CSA for the multifidus ( $P < 0.0001$ ), and in CSA ( $P < 0.001$ ) but not FSF ( $P = 0.17$ ) for the erector spinae. There was a significant effect of age for both muscles in FSF only ( $P < 0.0001$ ). CSA, cross-sectional area; FSF, fat signal fraction.



**Figure 4.** Univariate regression plots for multifidus FSF in men (A) and women (B) and erector spinae (C, D). Regression lines from healthy individuals from Crawford *et al*<sup>25</sup> are superimposed using a dotted line for comparison. (\*) indicates  $P < 0.05$  between regression intercepts of healthy and pathological populations. A trend for a difference between the intercepts (#) was seen for both muscles in men ( $P = 0.06$ ), and between slopes (+) both muscles in women ( $P = 0.08$ ). FSF, fat signal fraction.

(SD) FSF values at the L4 level in a healthy population of 21.2 (9.3)% in females and 15.3 (5.3)% in males. The previously reported values for mean FSF percentages are substantially lower than in the current study, for which mean FSF percentages in the multifidus were 45.3 (11.8)% in men and 51.6 (10.7)% in women with pathology in the multifidus. The higher percentage of fatty infiltrate may be because of differences in fat fraction calculation methodology, whereas

Fischer *et al* calculated fat fraction based on a single voxel measurement placed in the center of the muscle, our calculations included the entire muscle region. Additionally, differences in definition of the muscular region of interest can also influence these percentages, as the FSF values from the healthy comparison cohort were calculated based on volumetric, not cross-sectional, ROIs with potentially different muscular border definitions. We chose the posterior border

TABLE 3. Regression Estimates for Normal and Pathological FSF Values with Age					
Multifidus	Normal (N = 80)		Pathological (N = 199)		P
	Slope	Intercept	Slope	Intercept	
Men	0.003 (0.001)		0.004 (0.008)		0.28
		0.056 (0.028)		0.205 (0.488)	0.06
Women	0.002 (0.001)		0.004 (0.006)		0.08
		0.134 (0.049)		0.269 (0.389)	0.03
Erector spinae					
Men	0.002 (0.001)		0.003 (0.009)		0.31
		0.061 (0.025)		0.220 (0.517)	0.05
Women	0.002 (0.001)		0.004 (0.006)		0.08
		0.110 (0.052)		0.205 (0.368)	0.10

Values are represented as mean (SD); normal values are based on personal correspondence with Crawford *et al*<sup>25</sup>.

for our ROI based on anatomical observations in normal healthy people that the multifidus muscle compartment is encapsulated posteriorly by the lumbosacral fascia, although there is no current consensus on standardized methodology for defining ROIs of these muscles.

### Contributions of Age to CSA and FSF

In the current study, the results indicate that across a broad age range, age contributes to changes in FSF, but not CSA, in individuals with lumbar spine pathology. This is consistent with Fortin *et al*<sup>27</sup> who reported no associations between CSA and age, and Crawford *et al*,<sup>25</sup> who reported no associations between muscle volume and age, but in conflict with a number of other studies.<sup>28-30</sup> In one study that investigated lumbar spine muscle volume and fatty infiltrate across a young and old group of healthy individuals of similar body weight, age explained 18% to 36% of the variance in multifidus and erector spinae FSF unilaterally using T1-weighted MRI pixel intensity analyses.<sup>31</sup> Interestingly, when variance results of the current study were compared to variances from the healthy cohort, age explained a larger variance in FSF in individuals with pathology than healthy individuals in women, but not in men, even when comparisons were limited to similar age ranges. This suggests that pathology may have some differential effect across sexes. Importantly, though age accounts for approximately 30% of the variance in FSF for individuals with lumbar spine pathology, absolute levels of fatty infiltrate are still substantially higher than healthy individuals, especially in women, indicating that the presence of spine pathology dramatically increases the amount of muscle loss. This may suggest that changes in muscle quality and size are a result of muscle degenerative processes related specifically to pathology, instead of simple disuse atrophy that is associated with aging. This has implications for rehabilitative management strategies in these individuals, as traditional exercise approaches have not been shown to reverse degenerative muscle changes.<sup>12,32</sup> Investigating the underlying mechanisms responsible for pathological versus age related changes may help uncouple these patterns, and are likely relevant in understanding the potential for functional differences in muscle between the populations.

### LIMITATIONS

There were several limitations in this study. First, data were collected from retrospective chart reviews, so additional meaningful patient demographics such as duration of symptoms and body mass index were not consistently available across all subjects. Additionally, the MR images were only analyzed from a slice at a single level in an effort to standardize CSA across individuals. However, this may not have been the location of pain or structural pathology in many of the patients, as degree of spinal stenosis or other degenerative changes were not quantified from the images. Additionally, a single slice image taken from the L4 vertebrae may not be generalizable to changes across the entire lumbar spine. However, previous literature suggests

that data from L4 are highly correlated with overall fat fraction of the entire lumbar spine (although these fat fractions were based on muscular volume, not CSA, at the L4 level),<sup>25</sup> and that pathological changes in the lumbar spine that are associated with a specific structural abnormality are often seen at the L4 to 5 or L5 to S1 levels,<sup>33,34</sup> making this a logical choice for a single image analysis location. Methodologically, FSF values are also affected by changes in signal intensity as a result of inflammation. Although no patients reported specific inflammatory diseases, mild lumbar inflammation may result in overestimation of FSF values. Additionally, information on weight was only included in the charts of a subset of patients, and may be an additional confounder to CSA and FSF given their associated changes with age. However, in a subset of individuals whose weight data were available (N = 164), there was still no significant effect of age on CSA when weight was accounted for in the model ( $P > 0.86$ ), and there were no differences in the magnitude of the age effect on FSF.

### CONCLUSION

This retrospective study examined the contribution of age to muscle size and quality in a large cohort men and women undergoing surgery for lumbar spine pathology. The results indicate that individuals with pathology demonstrate similar CSAs, but larger overall levels of fatty infiltrate, compared to healthy controls across all ages. Although CSA is greater in men than women, it does not decline with age, whereas fatty infiltrate increases with age and pathology in both muscles. In women, the rate of increase in fatty infiltrate with age tends to be greater than their healthy counterparts. The larger volume of fat likely reduces the functional capacity of these important stabilizing muscles in the lumbar spine. Further research is required to elucidate the underlying mechanisms of age versus pathology-related changes in muscle quality and their functional implications.

### ➤ Key Points

- ❑ Fatty Infiltration, but not muscle size, increases with age in individuals with lumbar spine pathology.
- ❑ Fatty infiltration is higher across all age levels in individuals with lumbar spine pathology compared to healthy controls.
- ❑ Increased rates of fatty infiltration with age occur in women with lumbar spine pathology compared to their healthy counterparts, but not in men.

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## References

1. Andersson GB. Epidemiological features of chronic low-back pain. *Lancet* 1999;354:581–5.
2. D'hooge R, Cagnie B, Crombez G, et al. Increased intramuscular fatty infiltration without differences in lumbar muscle cross-sectional area during remission of unilateral recurrent low back pain. *Man Ther* 2012;17:584–8.
3. Alaranta H, Tallroth K, Soukka A, et al. Fat content of lumbar extensor muscles and low back disability: a radiographic and clinical comparison. *J Spinal Disord* 1993;6:137–40.
4. Freeman MD, Woodham MA, Woodham AW. The role of the lumbar multifidus in chronic low back pain: a review. *PM R* 2010;2:142–6.
5. Delmonico MJ, Harris TB, Visser M, et al., Health Ai, and Body. Longitudinal study of muscle strength, quality, and adipose tissue infiltration. *Am J Clin Nutr* 2009;90:1579–85.
6. Lieber RL, Bodine-Fowler SC. Skeletal muscle mechanics: implications for rehabilitation. *Phys Ther* 1993;73:844–56.
7. Fortin M, Battié MC. Quantitative paraspinal muscle measurements: inter-software reliability and agreement using OsiriX and ImageJ. *Phys Ther* 2012;92:853–64.
8. Hebert JJ, Kjaer P, Fritz JM, et al. The relationship of lumbar multifidus muscle morphology to previous, current, and future low back pain: a 9-year population-based prospective cohort study. *Spine (Phila Pa 1976)* 2014;39:1417–25.
9. Danneels LA, Vanderstraeten GG, Cambier DC, et al. Effects of three different training modalities on the cross sectional area of the lumbar multifidus muscle in patients with chronic low back pain. *Br J Sports Med* 2001;35:186–91.
10. Hebert JJ, Marcus RL, Koppenhaver SL, et al. Postoperative rehabilitation following lumbar discectomy with quantification of trunk muscle morphology and function: a case report and review of the literature. *J Orthop Sports Phys Ther* 2010;40:402–12.
11. Hebert JJ, Fritz JM, Thackeray A, et al. Early multimodal rehabilitation following lumbar disc surgery: a randomised clinical trial comparing the effects of two exercise programmes on clinical outcome and lumbar multifidus muscle function. *Br J Sports Med* 2015;49:100–6.
12. Käser L, Mannion AF, Rhyner A, et al. Active therapy for chronic low back pain: part 2. Effects on paraspinal muscle cross-sectional area, fiber type size, and distribution. *Spine (Phila Pa 1976)* 2001;26:909–19.
13. Beneck GJ, Kulig K. Multifidus atrophy is localized and bilateral in active persons with chronic unilateral low back pain. *Arch Phys Med Rehabil* 2012;93:300–6.
14. Chen YY, Pao JL, Liaw CK, et al. Image changes of paraspinal muscles and clinical correlations in patients with unilateral lumbar spinal stenosis. *Eur Spine J* 2014;23:999–1006.
15. Danneels LA, Vanderstraeten GG, Cambier DC, et al. CT imaging of trunk muscles in chronic low back pain patients and healthy control subjects. *Eur Spine J* 2000;9:266–72.
16. Hicks GE, Simonsick EM, Harris TB, et al. Cross-sectional associations between trunk muscle composition, back pain, and physical function in the health, aging and body composition study. *J Gerontol A Biol Sci Med Sci* 2005;60:882–7.
17. Kalichman L, Hodges P, Li L, et al. Changes in paraspinal muscles and their association with low back pain and spinal degeneration: CT study. *Eur Spine J* 2010;19:1136–44.
18. Le Cara EC, Marcus RL, Dempsey AR, et al. Morphology versus function: the relationship between lumbar multifidus intramuscular adipose tissue and muscle function among patients with low back pain. *Arch Phys Med Rehabil* 2014;95:1846–52.
19. Barry JJ, Lansdown DA, Cheung S, et al. The relationship between tear severity, fatty infiltration, and muscle atrophy in the supraspinatus. *J Shoulder Elbow Surg* 2013;22:18–25.
20. Raz Y, Henseler JF, Kolk A, et al. Patterns of age-associated degeneration differ in shoulder muscles. *Front Aging Neurosci* 2015;7:236.
21. Maden-Wilkinson TM, McPhee JS, Rittweger J, et al. Thigh muscle volume in relation to age, sex and femur volume. *Age (Dordr)* 2014;36:383–93.
22. Rosset A, Spadola L, Ratib O. OsiriX: an open-source software for navigating in multidimensional DICOM images. *J Digit Imaging* 2004;17:205–16.
23. Niemeläinen R, Briand MM, Battié MC. Substantial asymmetry in paraspinal muscle cross-sectional area in healthy adults questions its value as a marker of low back pain and pathology. *Spine (Phila Pa 1976)* 2011;36:2152–7.
24. Commean PK, Tuttle LJ, Hastings MK, et al. Magnetic resonance imaging measurement reproducibility for calf muscle and adipose tissue volume. *J Magn Reson Imaging* 2011;34:1285–94.
25. Crawford RJ, Filli L, Elliott JM, et al. Age- and level-dependence of fatty infiltration in lumbar paravertebral muscles of healthy volunteers. *AJNR Am J Neuroradiol* 2016;37:742–8.
26. Fischer MA, Nanz D, Shimakawa A, et al. Quantification of muscle fat in patients with low back pain: comparison of multi-echo MR imaging with single-voxel MR spectroscopy. *Radiology* 2013;266:555–63.
27. Fortin M, Yuan Y, Battié MC. Factors associated with paraspinal muscle asymmetry in size and composition in a general population sample of men. *Phys Ther* 2013;93:1540–50.
28. Gibbons LE, Videman T, Battié MC, et al. Determinants of paraspinal muscle cross-sectional area in male monozygotic twins. *Phys Ther* 1998;78:602–10.
29. Mannion AF, Käser L, Weber E, et al. Influence of age and duration of symptoms on fibre type distribution and size of the back muscles in chronic low back pain patients. *Eur Spine J* 2000;9:273–81.
30. McLoughlin RF, D'Arcy EM, Brittain MM, et al. The significance of fat and muscle areas in the lumbar paraspinal space: a CT study. *J Comput Assist Tomogr* 1994;18:275–8.
31. Valentin S, Licka T, Elliott J. Age and side-related morphometric MRI evaluation of trunk muscles in people without back pain. *Man Ther* 2015;20:90–5.
32. Airaksinen O, Herno A, Kaukanen E, et al. Density of lumbar muscles 4 years after decompressive spinal surgery. *Eur Spine J* 1996;5:193–7.
33. Magee DJ. Orthopedic physical assessment. Sciences EH, editor 2014.
34. Albert HB, Briggs AM, Kent P, et al. The prevalence of MRI-defined spinal pathoanatomies and their association with modic changes in individuals seeking care for low back pain. *Eur Spine J* 2011;20:1355–62.